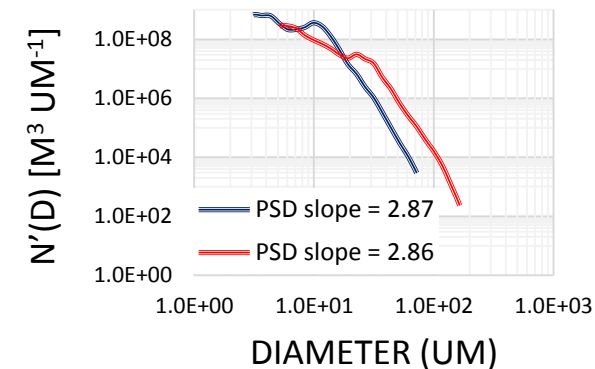
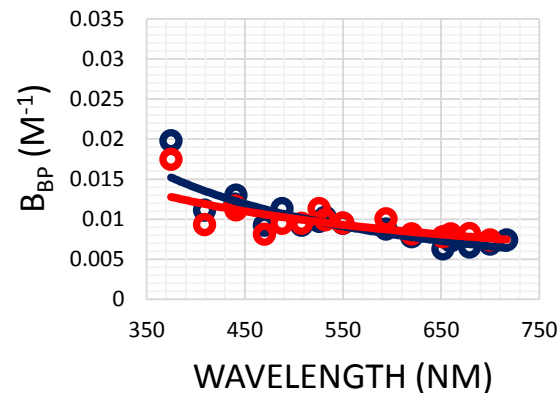
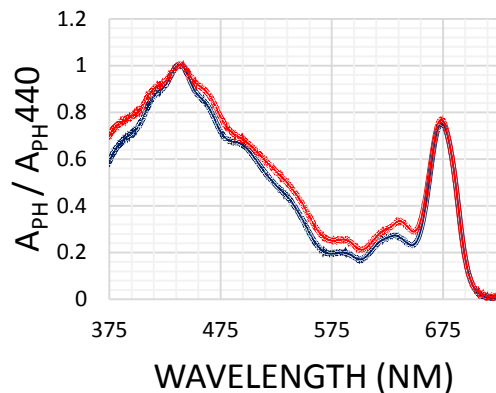
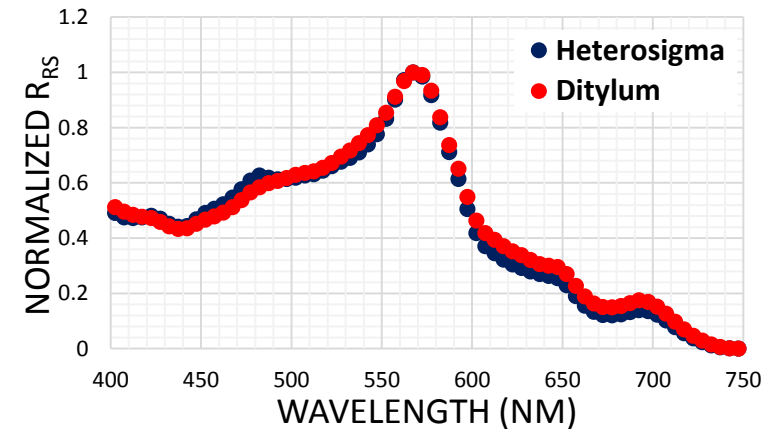
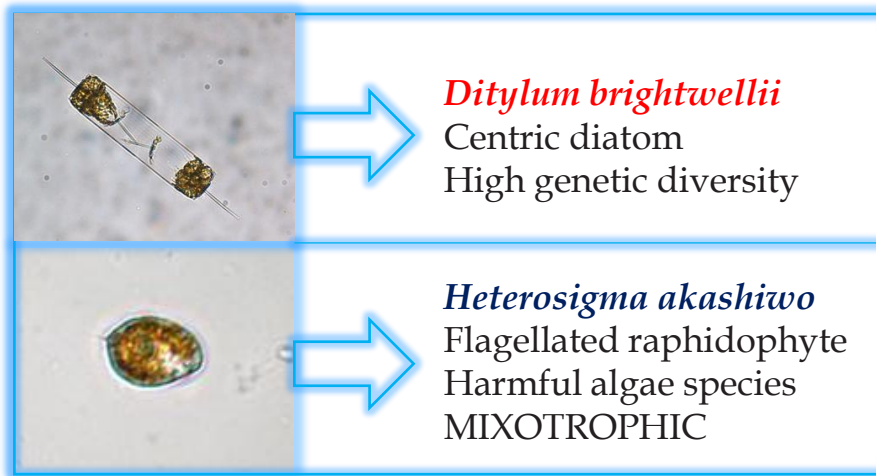
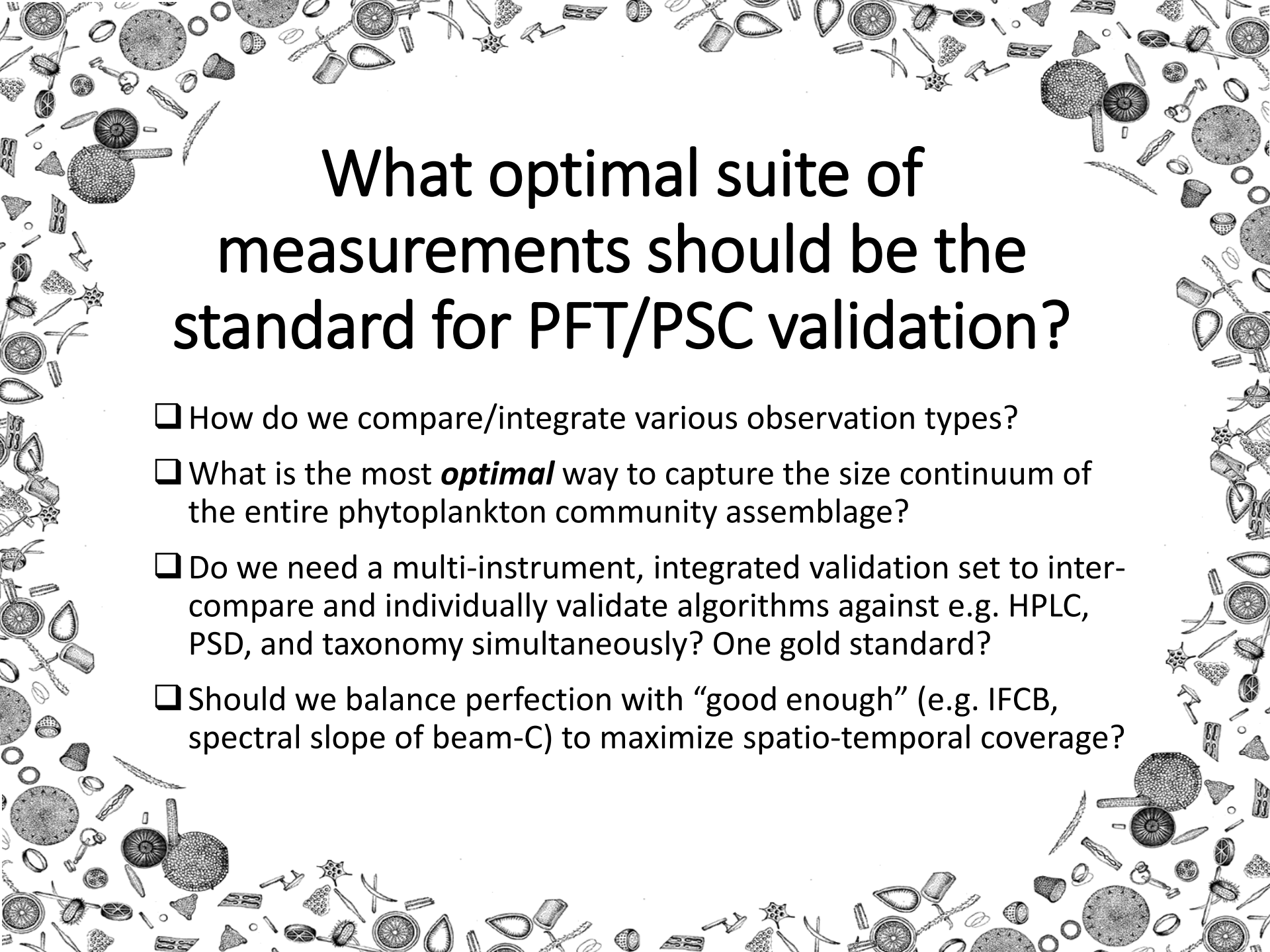


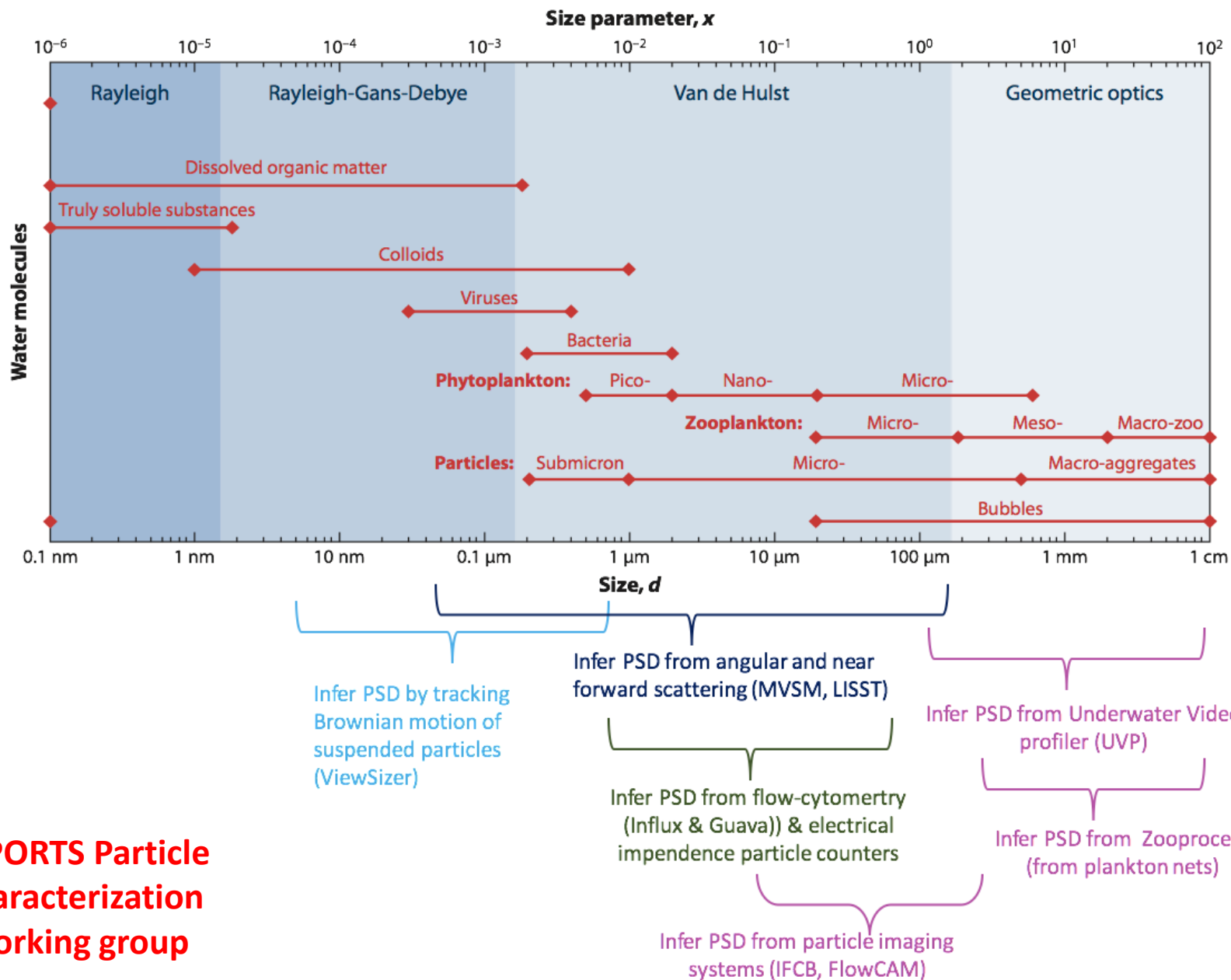
Challenges for PFT algorithm development/validation



A decorative border composed of numerous small, detailed black-and-white illustrations of various phytoplankton species, including diatoms, radiolarians, and other microorganisms, arranged in a circular pattern around the central text.

What optimal suite of measurements should be the standard for PFT/PSC validation?

- ☐ How do we compare/integrate various observation types?
- ☐ What is the most ***optimal*** way to capture the size continuum of the entire phytoplankton community assemblage?
- ☐ Do we need a multi-instrument, integrated validation set to inter-compare and individually validate algorithms against e.g. HPLC, PSD, and taxonomy simultaneously? One gold standard?
- ☐ Should we balance perfection with “good enough” (e.g. IFCB, spectral slope of beam-C) to maximize spatio-temporal coverage?



**EXPORTS Particle
Characterization
working group**

Karp-Boss et al. 2019

A decorative border composed of numerous small, detailed illustrations of various fruits and vegetables, including apples, oranges, lemons, limes, pears, and leafy greens, arranged in a circular pattern around the central text.

What types of ancillary data can augment PFT algorithm development/validation?

- ☐ e.g. BGC provinces, T-S diagrams, seasonal distribution, genomics?
- ☐ Should these be incorporated in algorithms?
- ☐ Can they be remotely sensed?
- ☐ What other sources of ancillary data will be useful for algorithm development and validation?